## ORIGINAL RESEARCH



# Dietary zinc requirement of Siberian sturgeon (*Acipenser baerii*, Brandt 1869) juveniles, based on the growth performance and blood parameters

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**Abstract** This study investigated the effect of dietary Zn levels on growth performance, feed utilization, and hematological parameters of juvenile Siberian sturgeon (*Acipenser baerii*, Brandt 1869). The five semi-purified diets were formulated by adding 0, 5, 10, 20, and 40 mg Zn kg<sup>-1</sup> diet in the form of zinc sulfate (ZnSO<sub>4</sub>) to provide the actual dietary value of 14.7, 20.8, 27.3, 37.7, and 46.4 Zn kg<sup>-1</sup> diet, respectively. The results showed that dietary Zn elevating level significantly improved the growth and feed utilization of the fish (p < 0.05), but not the hepatosomatic index (p > 0.05). Erythrocyte number, hemoglobin concentration, and hematocrit were increased with raising dietary Zn level (p < 0.05), whereas a regular trend was not observed in blood indices, including MCV, MCH, and MCHC. A significant increase was also found in the leucocyte number followed by their differential counts, except for eosinophil percentage. The broken line regression model indicated optimal dietary Zn level for growth maximization of fish was estimated to be 29.15 mg Zn kg<sup>-1</sup> diet. In spite of well correlation with dietary Zn level, no break point was observed to estimate the juvenile Siberian sturgeon requirement based on the erythrocyte numbers.

Keywords Zinc · Erythrocyte · Leucocyte · Blood parameters · Growth performance · Acipenser baerii

# **Abbreviations**

FW Final weight

SGR Specific growth rate
FCR Food conversion ratio
CF Condition factor
HSI Hepatosomatic index
RBC Red blood cell
WBC White blood cell

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Hb Hemoglobin HCT Hematocrit

MCV Mean corpuscular volume MCH Mean cell hemoglobin

MCHC Mean corpuscular hemoglobin concentration

#### Introduction

Zinc (Zn) is an essential micro-mineral required for various metabolic pathways, including growth, protein synthesis, energy metabolism, and immunity of animals, including fish (Houng-Yung et al. 2014; Lin et al. 2013). Zinc also serves as a functional cofactor for more than 200 metalloenzymes, such as superoxide dismutase, RNA polymerase, alkaline phosphatase, alcohol dehydrogenase, and carbonic anhydrase (Kucukbay et al. 2006; Fountoulaki et al. 2010; Luo et al. 2011; Liang et al. 2012; Lin et al. 2013). The importance of Zn in the antioxidant protection has been also illustrated in many aquatic organisms (Huang et al. 2015; Feng et al. 2011; Trevisan et al. 2014).

Although fish can uptake the Zn directly from the surrounding water via the gills, ambient waterborne Zn concentration in most freshwaters is known to be suboptimal to meet their metabolic requirements and a dietary supplementation is necessary to compensate for low dietborne Zn concentration (Gatlin and Phillips 1989; Fountoulaki et al. 2010; Luo et al. 2011). On the other hand, excessive Zn in the diet has been reported to compete with the absorption of other bivalent elements, including copper (Cu), iron (Fe), calcium (Ca), and cadmium (Cd) (Clearwater et al. 2002). Quantification of environmental quality parameters provides only limited information on the substances present in the environment aquatic and gives no information on the relationship between contaminant exposure and biological effects in aquatic organisms; therefore, the impact of pollutants by biomarkers becomes of relevant interest (Fazio et al. 2012, 2013; Chromcova et al. 2015). High Zn ingestion also reduces growth and feed utilization as the cause of appetite loss in Nile tilapia, *Oreochromis niloticus* (Do Carmo et al. 2004), Jian carp, *Cyprinus carpio* var. Jian (Tan et al. 2011), and juvenile yellow catfish, *Pelteobagrus fulvidraco* (Luo et al. 2011). Hence, dietary Zn level should be in an optimal range which can decrease the feed cost and prevent mineral leaching in aquatic environments (Buentello et al. 2009; Huang et al. 2015).

The dramatic reduction in sturgeon populations due to overfishing, poaching, pollution, and habitat alteration has led to the inevitability of commercial aquaculture as a tool for decreasing pressure on wild fish stocks as well as caviar and meat production (Beamesderfer and Farr 1997; Waldman and Wirgin 1997; Williot et al. 2002; Chebanov et al. 2002). The Siberian sturgeon (*Acipenser baerii*) is a one of the freshwater fish species which natively distributed in the major Siberian Rivers draining to the Kara, Laptev, and East Siberian seas (Bronzi et al. 2011; Wei et al. 2011). Fast growth rate during early life stages and strong adaptability properties to various climatic zones made the Siberian sturgeon a suitable candidate for aquaculture throughout the world (Hamlin et al. 2006; Abdolahnejad et al. 2015).

To date, numerous researches have been aimed to quantify dietary Zn requirement in fish, including 26–29 mg kg<sup>-1</sup> diet for hybrid tilapia, *O. niloticus* × *O. aureus* (Lin et al. 2008), 28.9–33.7 mg kg<sup>-1</sup> diet for juvenile grouper, *Epinephelus malabaricus* (Houng-Yung et al. 2014), 48.7 mg kg<sup>-1</sup> diet for juvenile Jian carp, *C. carpio* var. Jian carp (Tan et al. 2011), and 55.1 mg kg<sup>-1</sup> diet for juvenile grass carp, *Ctenopharyngodon idella* (Liang et al. 2012). Most of these recommendations have been established based on the growth performance, feed utilization, tissue concentration, and enzymatic activities. However, scare information is available regarding the Zn requirement for improvement of blood parameters in fish species under culture conditions. Accordingly, the aim of the present study was to evaluate the effects of elevated zinc levels on growth performance, feed utilization, and hematological parameters of juvenile Siberian sturgeon (*Acipenser baerii*, Brandt 1869).



#### Materials and methods

# Experimental diets

The formulation and proximate composition analysis of the basal diet is presented in Table 1. Casein, gelatin, and fish meal were used as dietary protein sources (51.0 g crude protein  $kg^{-1}$  diet). Corn oil and Kilka fish oil were used as the main dietary lipid source (11.6 g crude lipid  $kg^{-1}$  diet). Wheat flour, dextrin, and corn starch were also used as the carbohydrate source (26.0 g crude carbohydrate  $kg^{-1}$  diet).

The five isonitrogenous, isolipidic, and isoenergetic semi-purified diets were formulated to contain increasing levels of supplemental zinc  $(0, 5, 10, 20, \text{ and } 40 \text{ mg Zn kg}^{-1})$  in the form of hydrated zinc sulfate  $(\text{ZnSO}_4 \cdot 5\text{H}_2\text{O})$  at the expense of Alpha cellulose, respectively. Final dietary Zn concentrations were determined using a flame atomic absorption photometer (AA-6800; Shimadzu, Kyoto, Japan) after acid digestion equaling 14.7 (0), 20.8, 27.3, 37.7, and 46.4 mg Zn kg<sup>-1</sup> in the corresponding diets, respectively.

For the formulation of the experimental diets, all dry ingredients were weighed and mixed thoroughly for 30 min in a food mixer. Pre-blended premix of fish oil and corn oil was then added slowly while mixing of the ingredients continued for another 30 min. Afterward,  $ZnSO_4 \cdot 5H_2O$  was dissolved in double distilled water and mixed well with the other ingredients to form wet dough. The corresponsive of  $ZnSO_4 \cdot 5H_2O$  corresponding to each diet was dissolved in double distilled water and mixed well with the other diet ingredients until stiff dough resulted. The dough was then passed through a pelletizer having a die of 2 mm diameter and dried at 30 °C in drier for 24 h (Hidalgo et al. 2002). The pellets were then packed, sealed, and stored at -20 °C until used.

## Fish and experimental conditions

The bioassay was done in the Shahid Dr. Beheshti Reproduction and Rearing Center, Rasht (SDBRRC), Iran. At the beginning of the experiment, a total number of 300 Siberian sturgeon juveniles (*Acipenser baerii*), with initial body weight of  $26.52 \pm 0.94$  g (mean  $\pm$  SD), were separately stocked in group of 20 uniform-sized

Table 1 Diet formulation and proximate analysis of the basal diet for Siberian sturgeon (Acipenser baerii) juveniles

•	. •
Ingredients	g kg <sup>-1</sup> dry weight diet
Casein	380
Gelatin	80
Fish meal	50
Wheat flour	110
Dextrin	70
Corn starch	110
Corn oil	60
Fish oil	60
Vitamin premix <sup>1</sup>	2
Mineral premix <sup>2</sup>	1
Alpha cellulose	4
Chemical analysis	
Crude protein	510
Crude lipid	116
Crude carbohydrate	260

<sup>&</sup>lt;sup>1</sup> Vitamin premix (mg kg<sup>-1</sup> diet): thiamin, 15 mg; riboflavin, 30 mg; pyridoxine, 15 mg; cobalamin, 0.05 mg; niacin, 175 mg; folate, 5 mg; ascorbic acid, 500 mg; inositol, 1000 mg; biotin, 2.5 mg; calcium pantothenate, 50 mg; choline chloride, 2000 mg; DL-alpha tocopherol acetate, 60 IU; DL-cholecalciferol, 3000 IU; and cellulose was used as the carrier

 $<sup>^2</sup>$  Mineral premix (Zn free; g kg $^{-1}$ ): calcium carbonate (40% Ca), 2.15 g; magnesium oxide (60% Mg), 1.24 g; ferric citrate, 0.2 g; potassium iodide (75% I), 0.4 mg; cupric sulfate, 300 mg; manganese sulfate (33% Mn), 0.3 g; dibasic calcium phosphate (20% Ca, 18% P), 5 g; cobalt sulfate, 2 mg; sodium selenite (30% Se), 3 mg; potassium chloride, 0.9 g; sodium chloride, 0.4 g; and cellulose was used as the carrier



specimens to fifteen 500-L fiberglass tanks each containing 300 L freshwater. Each experimental diet was assigned to three tanks in a completely randomized design. Fish were acclimatized to the experimental condition for 2 weeks prior to the feeding trial and hand-fed with the basal diet (without Zn supplementation) to apparent satiation for Zn-depletion from the body reserves. Filtered water of the Sefidroud River was flowed through each tank to replace the whole tank water every 12 h. All fish groups were fed on the corresponding diets to apparent satiation two times a day for a period of 8 weeks. Fish consumed the respective diet in less than 2 min, and therefore, leaching of Zn to the rearing water was negligible. Tanks were carefully cleaned every other week to minimize algae and fungal growth at the same time with fish weaning for adjustment of daily feeding ratio.

During the acclimatization and experimental periods, the water conditions were kept at the appropriate range for Siberian sturgeon maintenance (average water temperature  $14.5 \pm 0.4$  °C, oxygen concentration  $7.43 \pm 0.25$  mg L<sup>-1</sup>, pH  $7.6 \pm 0.2$ , alkalinity  $247.5 \pm 3.8$  mg L<sup>-1</sup>, total hardness  $320.3 \pm 14.3$  mg L<sup>-1</sup>, and NH<sub>4</sub>  $0.14 \pm 0.01$  mg L<sup>-1</sup>). Natural photoperiod (approximately 10:14 light/dark) was maintained during the experimental period. Throughout the experimental period, the Zn concentration in the river water ranged from 0.0073 to 0.0217 µg Zn L<sup>-1</sup> without significant difference between the tank water (p < 0.05). All physico-chemical analysis were performed with DR 1900 portable spectrophotometer (Hach, Loveland, CO, USA) except for pH and Temperature which had been measured using portable AP110 pH Meter (Fisher Scientific, Pittsburgh, PA, USA) and temperature meter (YSI incorporated, Yellow Springs, OH, USA).

## Sample collection

Fish in each experimental tank were anesthetized with clove oil, *Syzygium aromaticum* (0.070 ppm; Kouřil et al. 2003) after 24 h starvation, and their body weight (g) and fork length (cm) were individually measured with a digital balance (Kern, Germany) and tapeline, respectively. The growth performance of Siberian sturgeon juveniles was determined by calculating specific growth rate (SGR), feed conversion ratio (FCR), protein efficiency ratio (PER), and hepatosomatic index (HSI) based on the standard formulae as follows (Luo et al. 2010):

Specific growth rate (SGR;  $\% d^{-1}$ ) = [Ln final weight (g) – Ln initial weight]/days  $\times 100$ 

Food conversion ratio (FCR) = feed consumption (g)/body weight gain (g)  $\times$  100

Condition factor (CF; g cm<sup>-3</sup>) = body weight (g)/fork length (cm<sup>3</sup>)  $\times$  100

Hepatosomatic index (HSI; %) = liver weight (g)  $\times$  100/fish weight (g)

Five fish from each experimental replicate were randomly selected to collect the blood sample from the caudal vessels. The fish blood was then pooled together and heparinized for hematological studies. A drop of each fish's blood was also used to make blood smear for leucocyte differential count.

# Hematological analysis

Counts of erythrocytes and leucocytes were performed based on the procedure suggested by Svobodova et al (1991) by mixing 20  $\mu$ L sample from each blood portion with 3980  $\mu$ L diluting fluids of red blood cells (RBCs) and white blood cells (WBCs), respectively. Cell counting was done under light microscope at magnification  $100\times$  using a Neubauer's counting chamber. Differential leucocyte counts were made from Leishman/Giemsa stained blood smears for 30 min, after drying at ambient temperature and fixing in 96% ethanol for 30 min (Klontz 1994). Microscopic examination was carried out under compound microscope at magnification  $400\times$  to count at least 100 leukocytes (Khoshbavar-Rostami et al. 2006).

The blood hemoglobin (Hb) concentration was determined based on the cyanmethemoglobin method using Drabkin's fluid (Drabkin 1950). An optical density (OD) was measured at 540 nm with a spectrophotometer and the hemoglobin level of blood was calculated as g dL<sup>-1</sup> by comparing with the standard cyanmethemoglobin based on the following formula:

$$Hemoglobin = \frac{OD_{test}}{OD_{standard}} \times 0.251.$$



Hematocrit was also calculated based on the microhematocrit method of Snieszko (1960) by centrifuging capillary tubes for 5 min at  $6000 \times g$  with a microcentrifuge NF 048. The derived hematological indices of mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), and mean corpuscular hemoglobin concentration (MCHC) were calculated using the standard formulae (Seiverd 1964).

# Statistical analysis

Levene's test was used to determine the homogeneity of variance, while the normality examined using the Kolmogorov–Smirnov test. The experimental parameters were subjected to one-way analysis of variance (one-way ANOVA), and comparisons between means were made by Tukey as a post-hoc test using the SPSS software (version 17). All data in the text are presented as mean  $\pm$  SD and statistical significance was accepted at the p < 0.05 level. The broken line model was also applied according to the procedure of Robbins (1986) to estimate the Zn requirement of Siberian sturgeon.

## **Results**

#### Growth performance

No mortality and abnormal behavior were observed in dietary treatments of Siberian sturgeon juveniles during the experimental period. The growth performance data are presented in Table 2.

The results indicated that dietary Zn level affected the growth performance of fish as final weight, and SGR and HSI values increased following the Zn level enhancement in experimental diets. The highest final weight was observed in 37.7 mg kg<sup>-1</sup> which differed significantly with control (p < 0.05). The SGR showed a similar pattern of FW with the peak of  $1.56 \pm 0.04\%$  d<sup>-1</sup> in the fish fed with 37.7 mg kg<sup>-1</sup> dietary Zn level, beyond that it was not significantly differed.

On the other hand, FCR significantly declined with increasing dietary Zn supplementation with the least of  $0.98 \pm 0.12$  in fish fed the diet containing 37.7 mg Zn kg<sup>-1</sup> (p < 0.05). The CF deceased by increasing dietary Zn level up to 37.7 mg kg<sup>-1</sup> diet and then increase with further supplementation. The CF was significantly decreased with increasing dietary Zn supplementation level, while no significant difference was found in the HSI of fish among the experimental treatments (p > 0.05).

Table 2 Growth performance and feed utilization of Siberian sturgeon (Acipenser baerii) juveniles, fed diets supplemented with graded zinc (Zn)

Dietary Zn level	FW (g)	SGR (% d <sup>-1</sup> )	FCR	CF (g cm <sup>-1</sup> )	HSI (%)
14.7 mg Zn	$50.86^{1a} \pm 2.65$	$1.14^{a} \pm 0.03$	$2.27 \pm 0.18$	$0.74^{\circ} \pm 0.01$	$2.39 \pm 0.05$
20.8 mg Zn	$53.07^{a} \pm 3.52$	$1.26^{a}\pm0.08$	$1.95 \pm 0.19$	$0.47^{\rm b} \pm 4.10$	$2.44 \pm 0.02$
27.3 mg Zn	$64.07^{\rm b} \pm 3.92$	$1.51^{\rm b} \pm 0.09$	$1.14 \pm 0.16$	$0.26^a \pm 0.03$	$2.67 \pm 0.05$
37.7 mg Zn	$65.34^{b} \pm 2.38$	$1.56^{b} \pm 0.04$	$0.98 \pm 0.12$	$0.24^a \pm 0.05$	$3.03 \pm 0.03$
46.4 mg Zn	$61.74^{ab} \pm 1.41$	$1.47^{ab} \pm 0.05$	$1.23 \pm 0.10$	$0.45^{\rm b} \pm 0.05$	$3.81 \pm 0.04$

Similar superscripts in each column show no significant difference between means for each treatment (p > 0.05)

Final weight (FW, g)

Specific growth rate (SGR,  $\% d^{-1}$ ) = [Ln final weight (g) – Ln initial weight]/days  $\times 100$ 

Food conversion ratio (FCR) = feed consumption (g)/body weight gain (g)

Condition factor (CF, g cm<sup>-3</sup>) = body weight (g)/fork length (cm<sup>3</sup>)  $\times$  100

Hepatosomatic index (HSI, %) = liver weight (g)  $\times$  100/fish weight (g)

<sup>&</sup>lt;sup>1</sup> Mean  $\pm$  SD of three groups of fish (n=3) with 20 fish per group except for HSI with 5 fish per group (n=20, except for HSI which n=5)



# Hematological parameters

Erythrocyte numbers of Siberian sturgeon juveniles were significantly (p < 0.05) enhanced with increasing dietary Zn supplementation from 14.7 to 37.7 mg kg<sup>-1</sup> diet, while did not significantly changed thereafter (Table 3). A similar pattern was observed for Hb and hematocrit, and the highest levels were seen in fish fed 46.4 mg Zn kg<sup>-1</sup>. In contrast, MCV and MCH declined with increasing dietary Zn level with the minimum level in fish fed with the diet containing 37.7 mg Zn kg<sup>-1</sup> which shows the value of 507.33  $\pm$  3.48 fL and 104.33  $\pm$  1.79 pg, respectively. However, no significant difference was found in the MCHC of fish fed with the supplemental Zn compared with those fed with the basal diet.

A significant (p < 0.05) increase was observed in leucocyte number of fish fed with the experimental diets (Table 4). Fish fed with control diet showed the least leucocyte count, while an increasing trend was recorded by increasing dietary Zn supplementation. All dietary Zn treatments induced significant increase in neutrophil and monocyte count of fish compared to the control treatment (p < 0.05). Conversely, the lymphocyte was significantly declined with increasing Zn supplementation to the basal diet (p < 0.05). The eosinophil percentage ranged between  $1.00 \pm 0.76$  and  $1.50 \pm 1.00$  with no significant differences among the treatments after 8-week feeding trial (p > 0.05).

## Dietary Zn requirement

The broken line regression model indicated that optimal dietary Zn level for growth maximization of Siberian sturgeon juveniles was estimated to be  $29.15 \text{ mg Zn kg}^{-1}$  diet based on the final weight (Fig. 1). The count of erythrocytes of Siberian sturgeon was correlated y = 4.0735x + 349.27,  $R^2 = 0.9013$ ) with dietary Zn supplementation (Fig. 2). However, no break point was observed to estimate the Zn requirement of fish based on the erythrocyte numbers.

## Discussion

The findings of the current research demonstrated that Siberian sturgeon juveniles have Zn requirement which could just be maintained through dietary supplementation. The lowest final weight and SGR observed in the fish fed with the basal diet, while they significantly enhanced by increasing Zn supplementation to the diets. The broken line analysis based on the final weight showed a requirement of 29.15 mg kg<sup>-1</sup> diet. However, the dietary Zn requirement varies between other freshwater fish species, i.e., 20 mg kg<sup>-1</sup> for channel catfish, *Ictalurus punctatus* (Gatlin and Wilson 1983), 15–30 mg kg<sup>-1</sup> for rainbow trout, *Oncorhynchus mykiss* (Ogino and Yang 1978), 26–29 mg kg<sup>-1</sup> for tilapia, *O. niloticus* × *O. aureus* (Lin et al. 2008), 20–25 mg kg<sup>-1</sup> for red drum, *Sciaenops ocellatus* (Gatlin et al. 1991), and 48.7 mg kg<sup>-1</sup> for juvenile Jian carp, *C carpio* var. Jian (Tan et al. 2011).

Table 3 Erythrocyte characteristics of Siberian sturgeon (Acipenser baerii) juveniles, fed diets supplemented with graded zinc

Dietary Zn level	RBC (×10 <sup>3</sup> cell mm <sup>-3</sup> )	Hb (g dL <sup>-1</sup> )	HCT (%)	MCV (fl)	MCH (pg)	MCHC (g dL <sup>-1</sup> )
14.7 mg Zn	$400.00^{1a} \pm 1.34$	$4.50^a \pm 0.55$	$22.00^a \pm 1.00$	$549.67^{\circ} \pm 2.42$	$112.00^{\rm b} \pm 1.63$	$20.19 \pm 0.38$
20.8 mg Zn	$434.00^{\rm b} \pm 1.12$	$4.87^{ab} \pm 0.40$	$23.67^{ab} \pm 1.07$	$544.67^{bc} \pm 4.20$	$112.00^{b} \pm 0.00$	$20.73 \pm 0.44$
27.3 mg Zn	$467.67^{c} \pm 1.24$	$5.17^{\rm bc} \pm 0.46$	$25.00^{\rm b}$ c $\pm 1.00$	$534.00^{\mathrm{bc}} \pm 2.50$	$110.00^{\rm b} \pm 1.00$	$20.67 \pm 0.25$
37.7 mg Zn	$519.00^{d} \pm 0.51$	$5.43^{\rm cd} \pm 0.40$	$26.33^{\rm cd} \pm 0.76$	$507.33^a \pm 3.48$	$104.33^{a} \pm 1.79$	$20.48 \pm 0.13$
46.4 mg Zn	$523.67^{\rm d} \pm 1.09$	$5.73^{d} \pm 0.46$	$27.67^{\rm d} \pm 0.76$	$528.00^{b} \pm 2.24$	$109.00^{\rm b} \pm 1.63$	$20.07 \pm 0.17$

Similar superscripts in each column show no significant difference between means for each treatment (p > 0.05)

RBC red blood cell, Hb hemoglobin, HCT hematocrit, MCV mean corpuscular volume, MCH mean cell hemoglobin, MCHC mean corpuscular hemoglobin concentration

Mean  $\pm$  SD of three groups of fish (n = 3) with five fish per group





Table 4 Leucocyte characteristics of Siberian sturgeon (Acipenser baerii) juveniles, fed diets supplemented with graded zinc

	Leucocyte ( $\times 10^3 \text{ mm}^{-3}$ )	Neutrophils (%)	Lymphocytes (%)	Monocytes (%)	Eosinophil (%)
14.7 mg Zn	$7.97^{1a} \pm 0.01$	$20.33^a \pm 1.23$	$76.67^{\circ} \pm 1.23$	$2.33^{a} \pm 0.76$	$1.00 \pm 0.76$
20.8 mg Zn	$9.20^{\rm bc} \pm 0.02$	$24.00^{b} \pm 1.00$	$72.33^{b} \pm 1.07$	$3.67^{ab} \pm 0.76$	$1.33 \pm 0.76$
27.3 mg Zn	$10.10^{\rm b} \pm 0.02$	$26.67^{\circ} \pm 1.24$	$67.33^a \pm 1.23$	$5.00^{b} \pm 1.00$	$1.50 \pm 1.00$
37.7 mg Zn	$11.63^{\circ} \pm 0.03$	$29.67^{d} \pm 0.76$	$65.00^{a} \pm 1.32$	$4.00^{b} \pm 1.00$	$1.33 \pm 0.76$
46.4 mg Zn	$12.33^{\circ} \pm 0.04$	$31.00^{d} \pm 1.00$	$64.00^{a} \pm 1.00$	$4.67^{\rm b} \pm 0.76$	$1.00 \pm 0.76$

Similar superscripts in each column show no significant difference between means for each treatment (p > 0.05)

<sup>&</sup>lt;sup>1</sup> Mean  $\pm$  SD of three groups of fish (n = 3) with five fish per group

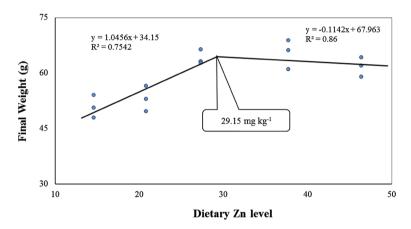


Fig. 1 Relationship between dietary Zn concentration and final weight in Siberian sturgeon (Acipenser baerii) juveniles, Acipenser baerii, based on the broken line regression analysis. Each point represents the mean of 20 fish in each replicate

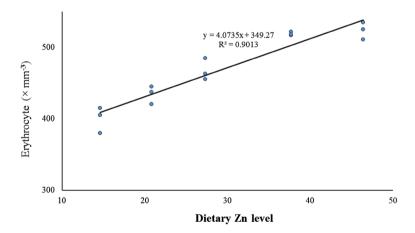


Fig. 2 Relationship between dietary Zn concentration and erythrocyte count in Siberian sturgeon (Acipenser baerii) juveniles, based on the regression analysis. Each point represents the mean of five fish in each replicate

Fish have similar cells types in their bloods with the same morphology and physiological functions found in mammals (Hrubec and Smith 2010). Although the normal ranges for biochemical key parameters have been identified for various raised fish species (Kopp et al. 2009; Xiaoyun et al. 2009; Satheeshkumar et al. 2011; Liang et al. 2012), there is scare information about the blood characteristics in Siberian sturgeon especially under culture conditions. Results of the present study illustrated that supplemental dietary Zn improves the hematological parameters of Siberian sturgeon. Erythrocyte counts were significantly increased by increasing Zn supplementation up to 37.7 mg kg<sup>-1</sup> and then plateau thereafter. Despite extensive researches on different metabolic pathways, there is little information about the effect of dietary Zn on erythrocytes synthesis in fish.



Zn is one of the essential elements affecting the protein synthesis in the erythrocytes, such as carbonic anhydrase and superoxide dismutase required for proper function of red blood cells (Hsieh et al. 2013; Hansch and Mendel 2009; Huang et al. 2015), which could explain the higher erythrocytes number of the fish fed diets with higher level of supplemental Zn in the present study.

Zn also plays an important role in hemoglobin synthesis by activating p-aminolevulinic acid (ALA) dehydrogenase, an enzyme essential for the porphobilinogen formation from two ALA molecules. In agreement with previous studies on other animals (El Hendy et al. 2001; Sobhanirad and Naserian 2012; Eze et al. 2015), results of the present research demonstrated that dietary Zn supplementation could significantly increase the Hb concentration in blood to  $5.73 \pm 0.46$  resulting higher ability of blood to transfer oxygen from the gills to the rest of body. Hb is an iron-containing metalloprotein in the red blood cells of all vertebrates which its synthesis is strongly restricted with the activity of enzymes involved in the Fe metabolism, such as ceruloplasmin, a ferroxidase enzyme associated with possible oxidation of ferrous iron to ferric iron (Lall 2002; Harrison and Arosio 1996). Although exact route of iron from its entry to the ferroxidase site is unknown, Zn offers the advantages which can replace Fe<sup>2+</sup> in iron-binding proteins without undergoing oxidation.

A controversial result was found in hematocrit of the fish fed different dietary Zn level compared to the other animals. Yadrick et al. (1989) expressed that supplemental dietary zinc decrease hematocrit, without changing the hemoglobin. In contrast, Bonham et al. (2003) stated that daily 40 mg Zn supplementation to the diet does not influence the blood parameters. In another investigation with 2-month-old rats, supplementation of 12 mg Zn kg<sup>-1</sup> diet decreased the hemoglobin 85% compared to the control animals (Zaporowska and Wasilewski 1992). Current results are similar by those of Do Carmo et al. (2004) that established an increase in hematocrit with augmentation of dietary Zn level reaching to the 27.67  $\pm$  0.76%, which show their higher capability of oxygen transfer from the gills to the tissues as well as more blood rheology and hemodynamics.

In the present study, MCV and MCH significantly decreased by increasing dietary Zn level from 14.7 to 37.7 mg kg<sup>-1</sup>, and then enhanced with further supplemental Zn to level of 46.4 mg kg<sup>-1</sup> diet. However, no significant difference was found in the MCHC of the fish fed the elevated dietary Zn levels. When looking at RBC count, hematocrit, MCV, an MCHC of Siberian sturgeon reared in the current research, it can be concluded that increase of hematocrit is just connected with increase in the number of erythrocytes without any significant change in their hemoglobin concentration individually. Hrubec and Smith (2010) stated that more active fish have higher hemoglobin level, smaller erythrocytes, and lower MCV. Therefore, smaller MCV of the Siberian sturgeon fed the higher dietary Zn levels illustrated their higher ability to stand against hypoxia which commonly happens in the culture condition.

Antioxidant activities and contribution in the defense system is one of the important functions of the Zn in fish (Powell 2000). Zinc deficiency provokes growth retardation, cataract, skin erosion, high mortality, and oxidative damage through the effects of free radical action (Powell et al. 1994; Salgueiro et al. 2000; Ogino and Yang 1978) and alters the status of antioxidant enzymes and substances (Prasad et al. 1993). The mechanism by which Zn exerts its antioxidant action is not well defined. However, it has been suggested that it increases the synthesis of metallothionein, a zinc-binding protein present in erythrocytes involved in various aspects of zinc metabolism, which acts as a free radical scavenger (Prasad et al. 1993; Bales et al. 1994). In the present research, the number of erythrocytes significantly increased in higher zinc concentrations (p < 0.05), reaching to more than 40% increase in fish fed the 46.4 mg kg<sup>-1</sup> diet compared to that in fish fed the basal diet.

Low dietary Zn concentration may characterize various diseases with aspects of an impaired immune response in fish representing more accurately reflects of tissue zinc. In vivo, Hu et al. (2001) observed that juvenile grouper had no difference in the non-specific immunity parameters (phagocytosis, complement, agglutination titer, and lysozyme activity) after feeding with zinc supplemented diets for 18 weeks. In vitro, zinc suppressed the lymphocyte proliferation of carp at a very low concentration (Cenini and Turner 1983; Ghanmi et al. 1989). Similarly, an 8-week feeding trial with Zn-supplemented diets had a significant effect on lymphocytes reduction which could be related to the fortification of innate immunity enhanced by the dietary Zn and less need for strengthening the humeral immunity.

On the other hand, result of the current study demonstrated an increase in the number of neutrophils as abundant type of granulocytes which form the essential part of innate immune system against pathogen infection. Previous studies illustrated that Zn may contribute to the host defense by interfering in the activation of neutrophils and subsequent oxidative burst (Finamore et al. 2008; Freitas et al. 2009), although the exact



role of this trace element, either as an activator or inhibitor, remains a matter of contention between the researchers.

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